Scripped on December 1, 203



The 50th anniversary of the Konopka and Benzer 1971 paper in PNAS: "Clock Mutants of *Drosophila melanogaster*"

Joseph S. Takahashi^{a,b,1}

Edited by Michael Rosbash, Howard Hughes Medical Institute, Waltham, MA, and approved August 3, 2021 (received for review July 10, 2021)

period gene | Konopka | Benzer | circadian | genetics

On September 1, 1971, unknowingly to most, the world changed for the fields of behavioral genetics and circadian clocks. Ronald Konopka, a graduate student with Seymour Benzer at Caltech, published a paper (1) that I would argue is the most important discovery ultimately leading to our current molecular understanding of the circadian clock in animals. In this classic paper, Ron and Seymour reported the isolation of three single-gene mutants in *Drosophila* that dramatically altered circadian rhythms in pupal eclosion and locomotor activity. One mutant exhibited no rhythmicity, another had a short 19-h period, and a third had a long 28-h period. Remarkably, all three mutants mapped to the same locus on the X chromosome. They named this gene *period*.

Seymour Benzer had recently joined the faculty at Caltech in 1967, after two previously successful careers in physics and molecular biology, and spurred on by Max Delbruck to do something more interesting, launched his third career in behavioral biology (2-5). Having done a sabbatical at Caltech with Roger Sperry, Seymour interacted with Ed Lewis, a giant in Drosophila genetics who trained with Alfred Sturtevant (descendent of Thomas Hunt Morgan) (6). Seymour chose *Drosophila* as a model system because its nervous system was intermediate in complexity "between a single neuron and the human brain" yet exhibited complex behavior and was amenable to genetic analysis (7). That same year, Seymour published his first paper (8) using mutagenesis and countercurrent technology to isolate phototaxis mutants in Drosophila. The opening sentence of this paper reads, "Complex as it is, much of the vast network of cellular functions has been successfully dissected, on a microscopic scale, by the use of mutants in which one element is altered at a time. A similar approach may be fruitful in tackling the complex structures and events underlying behavior, using behavioral mutations to indicate modifications of the nervous system."

Shortly afterward, Ron Konopka joined Seymour's laboratory as his first graduate student at Caltech (9, 10) (Fig. 1). Ron was previously interested in circadian rhythms, and with the tutelage of Ed Lewis, decided to do a screen for ethyl methanesulfonate-induced mutants on the X chromosome. Using the timing of eclosion of flies as a screen, Ron found an arrhythmic mutant in the first 200 lines that he screened. This led to what is known as Konopka's First Law: "If you don't find it among the first 200, quit" (9, 11). He went on to screen 1,900 lines and found two more mutants: the short period and long period mutants. Ron then went on to map the mutants using recombination and found them to be left of the white locus on X. To test for whether the three mutants were alleles of the same gene, he performed complementation tests for all combinations of the three mutants with each other and with wild type. None of the complementation tests produced a normal rhythm, suggesting the three mutants were alleles of the same gene. The arrhythmic and long mutants were recessive to wild type, and the short period mutant was semidominant. Crossing either the short or the long mutant over the arrhythmic mutant showed that

Author contribution: J.S.T. wrote the paper.

The authors declare no competing interest.

This article is a PNAS Direct Submission.

Published under the PNAS license.

See Classic companion article, "Clock mutants of Drosophila melanogaster," on page 2112 in issue 9 of volume 68.

See Classic Profile e2115546118, in vol. 118, issue 39.

¹Email: joseph.takahashi@utsouthwestern.edu.

This article contains supporting information online at https://www.pnas.org/lookup/suppl/doi:10.1073/pnas.2110171118/-/DCSupplemental. Published September 10, 2021.

^aHoward Hughes Medical Institute, University of Texas Southwestern Medical Center, Dallas, TX 75390; and ^bDepartment of Neuroscience, University of Texas Southwestern Medical Center, Dallas, TX 75390



Fig. 1. Photograph of Ron Konopka (Left) and Seymour Benzer (Right) taken in October 2000 in Pasadena, California. Photo courtesy of Howard Hughes Medical Institute, Holiday Lectures on Science, December 2000, "Clockwork Genes: Discoveries in Biological Time." Reprinted from ref. 14 by permission of SAGE Publications, Inc. copyright © 2004.

the arrhythmic mutant behaved as a null. Complementation tests with deficiencies of the X chromosome localized the gene within bands 3A6 to 3C2. Thus, Konopka and Benzer were able to infer that the arrhythmic mutant was a null mutant and that the short and long mutants were likely missense mutations. This prediction was confirmed many years later when the mutant alleles were sequenced (12, 13).

At the time that Konopka and Benzer published their paper, there was great skepticism that something as complex as behavior could be strongly influenced by single genes (4, 11). When Seymour reported the results to Max Delbruck, he told him it was impossible. To which Seymour responded, "But, Max, we found the gene, we've already done it!" Still Delbruck insisted that it was impossible and told Seymour, "I don't believe a word of it" (4, 11).

The Konopka and Benzer paper (1) completely refuted the notion that single genes could not influence behavior. Not only did the three period mutants affect the timing of eclosion, a developmental event, but the mutants affected the circadian locomotor rhythms of individual adult flies in the same way. Importantly, the phenotypes of the three alleles were so different: One could speed up the clock, one could slow it down, and one led to the abolition of circadian rhythms. These features emphasize the importance of choosing the best phenotypes to screen: For circadian rhythms, the period or rate of the underlying oscillator is most informative because there are few ways to change the rate of the oscillator in a nonspecific or irrelevant manner (14, 15).

This robust example of a single gene affecting circadian behavior reinforced Benzer's grand scheme that complex behavior could be dissected by Mendelian genetics and led to a groundswell of additional mutants in flies (7, 11). The Shaker mutant defined the first potassium channel at the molecular level (16, 17). The dunce mutant provided an entrée into the genetics of learning and memory (18, 19). The sevenless mutant defined a developmental pathway for photoreceptor determination (20-22). The bubblegum and methuselah mutants uncovered genes affecting neurodegeneration and life span (23, 24).

Despite the remarkable phenotypes of the period mutants, progress on the molecular nature of the period gene was slow. This required the development of recombinant DNA technology in the late 1970s and early 1980s in order to clone the gene. Thus, it would be another 13 y before the period gene would be identified at the molecular level independently by the laboratories of Jeff Hall and Michael Rosbash at Brandeis (25, 26) and by Mike Young at Rockefeller (27, 28). The cloning of per was not very informative at the time. There were few obvious protein motifs to infer function and a number of red herrings and dead ends were reported for the PER protein including its being a proteoglycan or a gap junction regulator (29-31). These side trips were eventually self-corrected by the Brandeis and Rockefeller laboratories (32, 33), and the focus reoriented toward gene expression and nuclear localization. It was shown that PER was actually a nuclear protein and initial antibody staining suggested that the PER protein expressed a circadian rhythm in the CNS and visual system of flies (34). Importantly, Hardin et al. (35) then showed that the per mRNA cycled in the brain of flies and that the per gene product could feedback on its own gene expression. This discovery led to the hypothesis of an autoregulatory loop involving PER negative feedback as the core of the circadian clock mechanism in *Drosophila*.

The magnitude of the *period* mutant discovery can measured in another way: The second core circadian gene to be discovered, *timeless*, did not occur until 1994, 23 y after *per* (36). Soon, it became clear that PER and TIM both cycled and formed a heterodimeric complex (37–40). The loop was finally closed with the discovery of the upstream regulators of *per* and *tim* by the transcription factors, CLOCK and CYCLE in 1998 (41–43).

From a personal perspective, the Konopka and Benzer (1971) paper was life changing for me. It was the exemplar for how to find circadian clock genes, and during midcareer, I switched from neuroscience to genetics because it became clear that in order to find clock genes in mammals, forward genetics was the only feasible path for discovering something that we knew nothing about (44). In 1994, we reported the isolation of the Clock mutant mouse, which was found in an N-ethyl-N-nitrosourea mutagenesis phenotypic screen for circadian locomotor rhythms (45). This mutant had an extreme circadian phenotype: a 28-h period and an eventual loss of sustained circadian rhythms in constant conditions. The mutant was semidominant and mapped to chromosome 5. Three years later, my laboratory was able to identify the gene using positional cloning and transgenic rescue approaches after 30 person years of effort (46, 47). The CLOCK protein was a bHLH-PAS transcription factor, and it acted in concert with another bHLH-PAS protein, BMAL1. Incredibly at the time, we found that CLOCK-BMAL1 could transactivate the *per* gene from *Drosophila* as well as the mouse *Per1* gene (48). Thus, CLOCK-BMAL1 linked directly to the original *period* gene and formed the basis for the previously missing activators of *per*. Identification of the *Drosophila* orthologs of *Clock* and *Bmal1*, *dClock*, and *cycle* (*dBmal1*), completed the description of the conserved core mechanism of the circadian clock in *Drosophila* and mammals (49). In the following two decades, remarkable progress was made in understanding the mechanism of circadian clocks in animals (50) and their relevance to health and medicine (51). Circadian clock proteins have direct molecular interactions with metabolism (52), immune function (53), cancer (54, 55), and neurodegeneration (56).

In 2017, Jeff Hall, Michael Rosbash, and Michael Young were awarded the Nobel Prize in Physiology or Medicine "for their discoveries of molecular mechanisms controlling the circadian rhythm" (33, 57). This would not have been possible without the truly groundbreaking paper by Konopka and Benzer (1) and the discovery of period. Sadly, neither Seymour nor Ron would witness the fruits of this paper leading to the Nobel Prize since they passed away in 2007 (2–4) and 2015 (9, 10), respectively. We owe a great debt to these two pioneering scientists who led the way in opening up the many black boxes underlying complex behavior.

See Movies S1 and S2 for interviews of Ronald Konopka and Seymour Benzer.

Data Availability. There are no data underlying this work.

- 1 R. J. Konopka, S. Benzer, Clock mutants of Drosophila melanogaster. Proc. Natl. Acad. Sci. U.S.A. 68, 2112-2116 (1971).
- 2 U. Banerjee, S. L. Zipursky, Seymour Benzer 1921–2007. Cell 131, 1217–1219 (2007).
- 3 N. M. Bonini, A tribute to Seymour Benzer, 1921–2007. Genetics 180, 1265–1273 (2008).
- 4 W. A. Harris, Seymour Benzer 1921–2007—The man who took us from genes to behaviour. PLoS Biol. 6, 205–207 (2008).
- 5 R. J. Greenspan, Seymour Benzer. 15 October 1921–30 November 2007. Biogr. Mem. Fellows R. Soc. 58, 23–32 (2012).
- 6 J. F. Crow, W. Bender, Edward B. Lewis, 1918–2004. Genetics 168, 1773–1783 (2004).
- **7** S. Benzer, From the gene to behavior. JAMA **218**, 1015–1022 (1971).
- 8 S. Benzer, Behavioral mutants of Drosophila isolated by countercurrent distribution. Proc. Natl. Acad. Sci. U.S.A. 58, 1112–1119 (1967).
- **9** J. C. Hall, Ronald J. Konopka. J. Biol. Rhythms **30**, 71–75 (2015).
- 10 M. Rosbash, Ronald J. Konopka (1947–2015). Cell 161, 187–188 (2015).
- 11 J. Wiener, Time, Love, Memory: A Great Biologist and His Quest for the Origins of Behavior (Knopf Press, New York, 1999).
- **12** M. K. Baylies, T. A. Bargiello, F. R. Jackson, M. W. Young, Changes in abundance or structure of the per gene product can alter periodicity of the *Drosophila clock*. *Nature* **326**, 390–392 (1987).
- **13** Q. Yu *et al.*, Molecular mapping of point mutations in the period gene that stop or speed up biological clocks in *Drosophila melanogaster. Proc. Natl. Acad. Sci. U.S.A.* **84**, 784–788 (1987).
- 14 J. S. Takahashi, Finding new clock components: Past and future. J. Biol. Rhythms 19, 339-347 (2004).
- 15 J. S. Takahashi, K. Shimomura, V. Kumar, Searching for genes underlying behavior: Lessons from circadian rhythms. Science 322, 909–912 (2008).
- 16 A. Kamb, L. E. Iverson, M. A. Tanouye, Molecular characterization of Shaker, a Drosophila gene that encodes a potassium channel. Cell 50, 405-413 (1987).
- 17 D. M. Papazian, T. L. Schwarz, B. L. Tempel, Y. N. Jan, L. Y. Jan, Cloning of genomic and complementary DNA from *Shaker*, a putative potassium channel gene from *Drosophila*. *Science* 237, 749–753 (1987).
- 18 Y. Dudai, Y. N. Jan, D. Byers, W. G. Quinn, S. Benzer, dunce, a mutant of Drosophila deficient in learning. Proc. Natl. Acad. Sci. U.S.A. 73, 1684–1688 (1976).
- 19 C. N. Chen, S. Denome, R. L. Davis, Molecular analysis of cDNA clones and the corresponding genomic coding sequences of the *Drosophila* dunce⁺ gene, the structural gene for cAMP phosphodiesterase. *Proc. Natl. Acad. Sci. U.S.A.* 83, 9313–9317 (1986).
- 20 W. A. Harris, W. S. Stark, J. A. Walker, Genetic dissection of the photoreceptor system in the compound eye of *Drosophila melanogaster*. J. Physiol. 256, 415–439 (1976).
- 21 U. Banerjee, P. J. Renfranz, J. A. Pollock, S. Benzer, Molecular characterization and expression of sevenless, a gene involved in neuronal pattern formation in the Drosophila eye. Cell 49, 281–291 (1987).
- 22 E. Hafen, K. Basler, J. E. Edstroem, G. M. Rubin, Sevenless, a cell-specific homeotic gene of *Drosophila*, encodes a putative transmembrane receptor with a tyrosine kinase domain. Science 236, 55–63 (1987).
- 23 Y. J. Lin, L. Seroude, S. Benzer, Extended life-span and stress resistance in the Drosophila mutant methuselah. Science 282, 943–946 (1998).
- 24 K. T. Min, S. Benzer, Preventing neurodegeneration in the Drosophila mutant bubblegum. Science 284, 1985–1988 (1999).
- 25 P. Reddy et al., Molecular analysis of the period locus in *Drosophila melanogaster* and identification of a transcript involved in biological rhythms. *Cell* 38, 701–710 (1984).
- 26 W. A. Zehring et al., P-element transformation with period locus DNA restores rhythmicity to mutant, arrhythmic *Drosophila melanogaster*. *Cell* 39, 369–376 (1984).
- 27 T. A. Bargiello, M. W. Young, Molecular genetics of a biological clock in Drosophila. Proc. Natl. Acad. Sci. U.S.A. 81, 2142-2146 (1984).
- 28 T. A. Bargiello, F. R. Jackson, M. W. Young, Restoration of circadian behavioural rhythms by gene transfer in Drosophila. Nature 312, 752-754 (1984).
- 29 F. R. Jackson, T. A. Bargiello, S. H. Yun, M. W. Young, Product of per locus of Drosophila shares homology with proteoglycans. Nature 320, 185–188 (1986).
- 30 P. Reddy, A. C. Jacquier, N. Abovich, G. Petersen, M. Rosbash, The period clock locus of D. melanogaster codes for a proteoglycan. Cell 46, 53-61 (1986).
- 31 T. A. Bargiello et al., The *Drosophila clock* gene per affects intercellular junctional communication. *Nature* 328, 686–691 (1987).

- 32 L. Saez et al., Per—No link to gap junctions. Nature 360, 542 (1992).
- 33 M. Rosbash, Circadian rhythms and the transcriptional feedback loop (Nobel Lecture). Angew. Chem. Int. Ed. Engl. 60, 8650-8666 (2021).
- 34 X. Liu et al., The period gene encodes a predominantly nuclear protein in adult Drosophila. J. Neurosci. 12, 2735–2744 (1992).
- **35** P. E. Hardin, J. C. Hall, M. Rosbash, Feedback of the *Drosophila period* gene product on circadian cycling of its messenger RNA levels. *Nature* **343**, 536–540 (1990).
- **36** A. Sehgal, J. L. Price, B. Man, M. W. Young, Loss of circadian behavioral rhythms and per RNA oscillations in the *Drosophila* mutant timeless. *Science* **263**, 1603–1606 (1994).
- 37 M. P. Myers, K. Wager-Smith, C. S. Wesley, M. W. Young, A. Sehgal, Positional cloning and sequence analysis of the *Drosophila clock* gene, *timeless. Science* 270, 805–808 (1995).
- 38 A. Sehgal et al., Rhythmic expression of timeless: A basis for promoting circadian cycles in period gene autoregulation. Science 270, 808-810 (1995).
- **39** N. Gekakis et al., Isolation of timeless by PER protein interaction: Defective interaction between timeless protein and long-period mutant PERL. Science **270**, 811–815 (1995).
- 40 H. Zeng, Z. Qian, M. P. Myers, M. Rosbash, A light-entrainment mechanism for the Drosophila circadian clock. Nature 380, 129–135 (1996).
- 41 R. Allada, N. E. White, W. V. So, J. C. Hall, M. Rosbash, A mutant *Drosophila* homolog of mammalian Clock disrupts circadian rhythms and transcription of *period* and *timeless. Cell* 93, 791–804 (1998).
- 42 J. E. Rutila et al., CYCLE is a second bHLH-PAS clock protein essential for circadian rhythmicity and transcription of *Drosophila period* and *timeless*. Cell 93, 805–814 (1998).
- 43 T. K. Darlington et al., Closing the circadian loop: CLOCK-induced transcription of its own inhibitors per and tim. Science 280, 1599-1603 (1998).
- 44 J. S. Takahashi, L. H. Pinto, M. H. Vitaterna, Forward and reverse genetic approaches to behavior in the mouse. Science 264, 1724–1733 (1994).
- 45 M. H. Vitaterna et al., Mutagenesis and mapping of a mouse gene, Clock, essential for circadian behavior. Science 264, 719-725 (1994).
- 46 D. P. King et al., Positional cloning of the mouse circadian clock gene. Cell 89, 641-653 (1997).
- 47 M. P. Antoch et al., Functional identification of the mouse circadian Clock gene by transgenic BAC rescue. Cell 89, 655-667 (1997).
- 48 N. Gekakis et al., Role of the CLOCK protein in the mammalian circadian mechanism. Science 280, 1564–1569 (1998).
- 49 R. Allada, P. Emery, J. S. Takahashi, M. Rosbash, Stopping time: The genetics of fly and mouse circadian clocks. Annu. Rev. Neurosci. 24, 1091-1119 (2001).
- 50 J. S. Takahashi, Transcriptional architecture of the mammalian circadian clock. Nat. Rev. Genet. 18, 164–179 (2017).
- 51 R. Allada, J. Bass, Circadian mechanisms in medicine. N. Engl. J. Med. 384, 550-561 (2021).
- 52 J. Bass, J. S. Takahashi, Circadian integration of metabolism and energetics. Science 330, 1349-1354 (2010).
- 53 C. Scheiermann, J. Gibbs, L. Ince, A. Loudon, Clocking in to immunity. Nat. Rev. Immunol. 18, 423-437 (2018).
- 54 S. Masri, P. Sassone-Corsi, The emerging link between cancer, metabolism, and circadian rhythms. Nat. Med. 24, 1795–1803 (2018).
- 55 F. Battaglin et al., Clocking cancer: The circadian clock as a target in cancer therapy. Oncogene 40, 3187–3200 (2021).
- 56 E. S. Musiek, D. M. Holtzman, Mechanisms linking circadian clocks, sleep, and neurodegeneration. Science 354, 1004–1008 (2016).
- 57 M. W. Young, Time travels: A 40-year journey from *Drosophila*'s clock mutants to human circadian disorders (Nobel Lecture). Angew. Chem. Int. Ed. Engl. 57, 11532–11539 (2018).

